



Proposition de projet de doctorat 2024-2027 – ED 406, Chimie moléculaire de Paris centre

## **Analysing interactions between antimicrobial peptides and peptidoglycan by affinity photocrosslinking coupled to MS**

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### **Project description:**

Antimicrobial peptides (AMPs) are a broad class of peptides with promising activity against bacteria or fungi and represent interesting starting points for the development of new therapeutic approaches to treat bacterial infections. Many AMPs act by interacting with bacterial lipid membranes and potentially cell-wall components such as peptidoglycan (PGN), although there are very few direct pieces of evidence of the latter.

We recently isolated DMS-DA6, an AMP showing a specific activity towards Gram-positive bacteria<sup>1</sup>. Gram-negative and Gram-positive cell walls differ greatly in terms of surface glycoconjugates. In the case of Gram-positive bacteria, a very thick layer of PGN and negatively charged lipoteichoic are found in the most outer part of the bacteria. PGN and LTA could act as peptide “sponges”, increasing their local concentration, or as peptide “traps”, keeping them away from the lipid membrane. We believe DMS-DA6 interacts with PGN but so far, we only have indirect evidence for this interaction.

Our objective is to obtain direct evidence of DMS-DA6/PGN interactions and identify the structural patterns involved in these interactions using affinity photocrosslinking coupled to MS on reconstituted models and live bacteria, and characterise these interactions by calorimetry. The long-term objective is to design new AMPs with optimised sequences.

We will use two complementary affinity photocrosslinking approaches in model systems or live bacteria. First, the peptide will carry a photolabel, that can easily be inserted in the sequence by solid-phase peptide synthesis. With such a design, we aim to identify glycoconjugate partners, based on the expertise we developed for the study of peptide/lipid interactions<sup>2-4</sup>. These peptides will be used in interaction with PGN extracted from bacteria, or directly on live bacteria once the analytical process is well established.

In parallel, we want to develop a new and original approach to study PGN/AMP interactions by metabolically introducing a photoreactive label in the stem peptide of PGN. This approach relies on biochemical engineering of PGN<sup>5</sup>, using either modified D-amino acids<sup>6,7</sup> or chemically modified Sortase A substrates<sup>8</sup>. Such approaches have previously been used to introduce molecular handles for post-labeling (alkyne, azide, thiols...) or fluorophores on live bacteria, but to our knowledge, it has never been used to introduce a photoreactive label.

This project should yield valuable information on AMP/PGN interactions. It will give insight on the general mechanism of membrane permeation by AMPs. It should help predict the activity of AMPs on

bacterial strains and enable us to rationalise the design of new AMPs with optimised sequences for better bacteria targeting.

This project is at the interface of chemistry and biochemistry and will involve a large set of techniques: peptide synthesis, mass spectrometry, cell culture, model membranes, etc...all available at the host laboratory (LBM). This PhD project will be supervised by Emmanuelle Sachon (HDR) and Astrid Walrant (HDR in preparation), whose expertise encompass mass spectrometry, affinity photolabelling, membrane active peptides and characterisation of biomolecular interactions.

#### References:

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- (2) Bechtella, L.; Kirschbaum, C.; Cosset, M.; Clodic, G.; Matheron, L.; Bolbach, G.; Sagan, S.; Walrant, A.; Sachon, E. Benzophenone Photoreactivity in a Lipid Bilayer To Probe Peptide/Membrane Interactions: Simple System, Complex Information. *Anal. Chem.* **2019**, *91* (14), 9102–9110.
- (3) Walrant, A.; Sachon, E. Photolabeling Strategies to Study Membranotropic Peptides Interacting with Lipids and Proteins in Membranes. *Bioconjug. Chem.* **2021**, *32* (8), 1503–1514.
- (4) Bechtella, L.; Chalouhi, E.; Milán Rodríguez, P.; Cosset, M.; Ravault, D.; Illien, F.; Sagan, S.; Carlier, L.; Lequin, O.; Fuchs, P. F. J.; et al. Structural Bases for the Involvement of Phosphatidylinositol-4,5-Bisphosphate in the Internalization of the Cell-Penetrating Peptide Penetratin. *ACS Chem. Biol.* **2022**, *17* (6), 1427–1439.
- (5) Siegrist, M. S.; Swarts, B. M.; Fox, D. M.; Lim, S. A.; Bertozzi, C. R. Illumination of Growth, Division and Secretion by Metabolic Labeling of the Bacterial Cell Surface. *FEMS Microbiol. Rev.* **2015**, *39* (2), 184–202.
- (6) Kuru, E.; Hughes, H. V.; Brown, P. J.; Hall, E.; Tekkam, S.; Cava, F.; De Pedro, M. A.; Brun, Y. V.; Vannieuwenhze, M. S. In Situ Probing of Newly Synthesized Peptidoglycan in Live Bacteria with Fluorescent D-Amino Acids. *Angew. Chemie - Int. Ed.* **2012**, *51* (50), 12519–12523.
- (7) Ferraro, N. J.; Kim, S.; Im, W.; Pires, M. M. Systematic Assessment of Accessibility to the Surface of *Staphylococcus Aureus*. *ACS Chem. Biol.* **2021**, *16* (11), 2527–2536.
- (8) Nelson, J. W.; Chamesian, A. G.; McEnaney, P. J.; Murelli, R. P.; Kazmiercak, B. I.; Spiegel, D. A. A Biosynthetic Strategy for Re-Engineering the *Staphylococcus Aureus* Cell Wall with Non-Native Small Molecules. *ACS Chem. Biol.* **2010**, *5* (12), 1147–1155.

#### Application process:

Applications are open from April 9<sup>th</sup> to May 5<sup>th</sup> and have to be deposited on ADUM:

<https://adum.fr/candidature/>

Applications must include a detailed CV, Master's grades (M1 and M2), one or two recommendation letters from direct supervisors (M1 and M2 internships) and a cover letter describing your motivations for the project.

Please also send your application directly to Emmanuelle Sachon and Astrid Walrant ([emmanuelle.sachon@sorbonne-universite.fr](mailto:emmanuelle.sachon@sorbonne-universite.fr) ; [astrid.walrant@sorbonne-universite.fr](mailto:astrid.walrant@sorbonne-universite.fr))